

# KeratinoSens®

**Assay Ready Cells** 

# Certificate of Analysis

Lot-N°: 92-191122JP03

## **BATCH SPECIFICATIONS**

Cell Designation: KeratinoSens® Source: Givaudan

Cat-N°: RE232 Lot-N°: 92-191122JP03

Packaging Unit: 2.5 million cells / vial Assay Medium: H

Passage: <25 Storage: below -150°C (e.g. liN<sub>2</sub>)

Approval Date: 12.12.2019 Approved by:

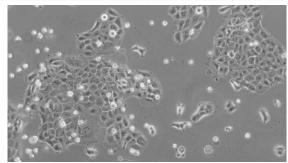
Expiry Date: 22.11.2021 Augu Ciura

Susan Ciura (Head of Quality Control)

## **QUALITY CONTROL**

	Batch Results	Specification Limits
Cell Count	2.36E+06	≥ 90 % of nominal cell count
Homogeneity (cell count)	97 %	≤ 90 %
Viability (after thawing)	97 %	≥ 90 %
Aggregation	1.2	≤ 2.0
Debris Ratio	0.2	≤ 1.0
Proliferative Capacity	100 %	≥ 70 %
Sterility (bacteria, yeast, fungi)	passed	sterile after 4 days
Sterility (mycoplasma)	passed	negative by PCR
Morphology	passed	unaltered to reference
EC <sub>1.5</sub> DNCB	2.07 μΜ	≤12.5 μM
EC <sub>1.5</sub> EGDMA	45.08 μM	30 - 100 μΜ
EC <sub>1.5</sub> Lactic Acid	> 2000 μM	≥ 1000 µM

## **MORPHOLOGY:**



Morphology of KeratinoSens Assay Ready Cells



#### **METHODS:**

Cell Viability Parameters: Viability parameters (viable cell count after thawing, grade of aggregation, percentage of debris)

were determined from a pooled sample (SOP-2015-02). Briefly, Assay Ready Cells were thawed in a water bath. 50  $\mu$ l of each sample were pooled, diluted 1:1000 in CASY Ton buffer and measured (3 replica) in a CASY TT automatic cell counter. Vial to vial variation was determined in a plate-based viability assay. Equal aliquots of each cell samples were dispensed into a 96 well-plate. A cellular dye was added to the well to obtain fluorescent signal which is proportional to

the viable cell count.

Proliferative Capacity: 1,0E+04 cells from the Assay Ready Cell samples and from a reference of a continuously passaged

culture were seed in a 96-well plate (8 replica each). After 48 hours of cultivation the proliferation of the cells was determined by addition of a metabolic cell dye (Resazurin). The proliferative capacity of the Assay Ready Cells is calculated in percent of the exponentially growing reference.

(SOP-2017-03).

Sterility Testing: Assay Ready Cells were seed in two specific bacteria growth brothes (Tryptic Soy Broth for aerob

and Thioglycollate broth for anaerob conditions) and cultivated over a course of 14 days. After day 1, 4, 7 and 14 the cultures were analyzed microscopically for cell growth, cell morphology, and incidences of contamination (bacteria, yeast, or fungi). For mycoplasma testing from a three days old, sub-confluent culture 500  $\mu$ l of the supernatant was taken and analyzed by PCR using a Mycoplasma detection kit (Minerva). The Assay was performed according to the manufacturer

protocol (SOP-2015-06).

Functional Testing: Assay Ready Cells were thawed, washed once in 10ml assay medium and seeded into a 96-well

plate at 10.000 cells/well. Incubation for 24 hours at 37 °C and 5 %  $CO_2$ . The supernatant was discarded after adherence of the cells and replaced by serial dilutions of the reference chemicals DNCB, EGDMA and Lactic Acid. After 48 h of incubation at 37 °C and 5 %  $CO_2$ ,  $50\mu$ l of OneGlo (Promega), a luciferase substrate was added to each well. After 20 min, the luminescence was

measured with an integration time of 1 s/well in a multiplate reader.

#### **LIMITED USE**

The product is sold under the terms of a Limited Use Label License attached to the product. By breaking the seal of the product package, the user explicitly agrees to the license terms. Assay Ready Cells are for immediate assay use only. The user shall not propagate, passage, or refreeze the cells.